

Total synthesis of (+)-camptothecin

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Abstract—A total synthesis of (+)-camptothecin is described. The approach is based on a room temperature LUMO_{diene}-controlled Diels–Alder cycloaddition of the electron deficient *N*-sulfonyl-1-aza-1,3-butadiene **6** with the electron rich dienophile 1,1,3,3-tetraethoxypropene (**7**) for assembly of a pyridine precursor to the D-ring pyridone. The 20(*S*) tertiary alcohol was installed through a Sharpless asymmetric dihydroxylation reaction on the methyl vinyl ether **12**, using the 3,4,5-trimethoxyphenyl-derived pyrimidine DHQ dimer ligand **16**. In a single reaction vessel, the C and E rings were closed using an acid-catalyzed deprotection of the benzylic ethers to afford the corresponding benzylic bromides (**18**) which underwent intramolecular nucleophilic displacement by the carboxylate and pyridone nitrogen to furnish (+)-camptothecin. © 2002 Elsevier Science Ltd. All rights reserved.

(+)-Camptothecin (CPT, **1**, Fig. 1) is a pentacyclic alkaloid isolated and characterized by Wall and co-workers in 1966 from *Camptotheca acuminata* and shown to exhibit potent cytotoxic activity against a range of tumor cell lines.^{1–3} Subsequent to its discovery, CPT was shown to stabilize the cleavable complex of double-stranded DNA and topo-

isomerase I.⁴ Topoisomerase I is responsible for the cleavage of double-stranded DNA to produce a covalently bound 3'-phosphate-enzyme complex⁵ and is enlisted to unwind supercoiled DNA ahead of the replication fork.⁶ Recent modeling studies of a ternary complex suggest camptothecin intercalates from the minor groove side of DNA forming key hydrogen bond interactions between its lactone and arginine 364 of human topoisomerase I.⁷ This prevents religation of double-stranded DNA, provides stabilization of the single strand break, and results in a permanent DNA lesion that leads to cell death.^{8–10} Although CPT itself is not used clinically for the treatment of cancer, two closely related analogues Topotecan (**2**) and Irinotecan (**3**) have proven effective in the clinic.^{11,12}

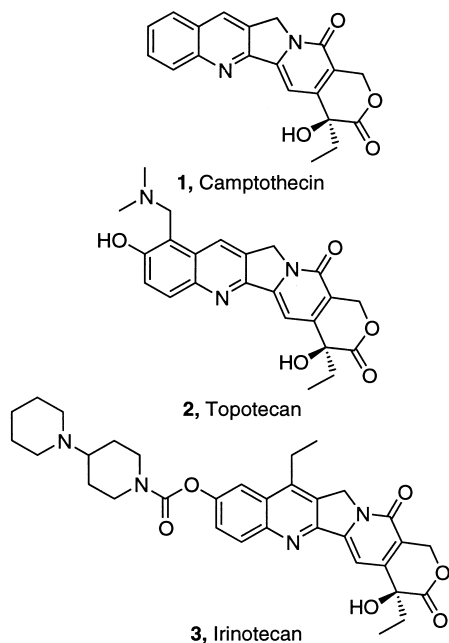


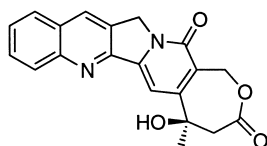
Figure 1.

Keywords: (+)-camptothecin; Diels–Alder cycloaddition; *N*-sulfonyl-1-aza-1,3-butadiene.

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Since its discovery, camptothecin has been the subject of extensive research especially for the design of water soluble derivatives^{11–18} and more recently, stable lactone analogues.^{19–23} At physiological pH, the lactone is easily hydrolyzed to the biologically inactive carboxylate, which binds to human serum albumin, lowering the effective concentration of CPT.^{24,25} This facile hydrolysis of the lactone represents a challenging problem and its labile nature has been attributed to the α -hydroxy group which is believed to accelerate the hydrolysis through intramolecular hydrogen bonding.²¹ Recently, the development of new analogues of CPT which are more stable has been pursued and culminated in the development of homocamptothecin (**4**, Fig. 2).^{19,20} Homocamptothecin contains a methylene spacer between the tertiary alcohol and the lactone carbonyl, resulting in prolonged activity in cellular assays.²¹

Although numerous analogues of CPT containing modifications in the lactone ring have been prepared, none have

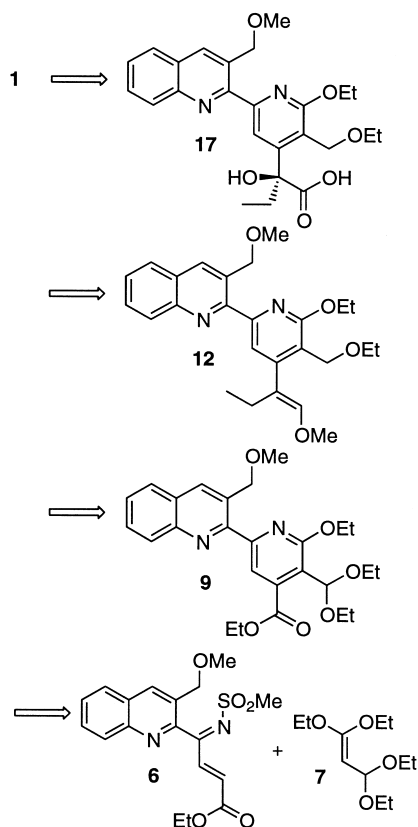


4, Homocamptothecin

Figure 2.

proven to be as active as CPT itself. Herein we detail the development of a versatile route for the asymmetric synthesis of CPT,²⁶ which allows for modification of the E-ring permitting access to new analogues that may help to define the key interactions between CPT/DNA/topoisomerase I and ultimately aid in the development of second generation CPT derivatives.

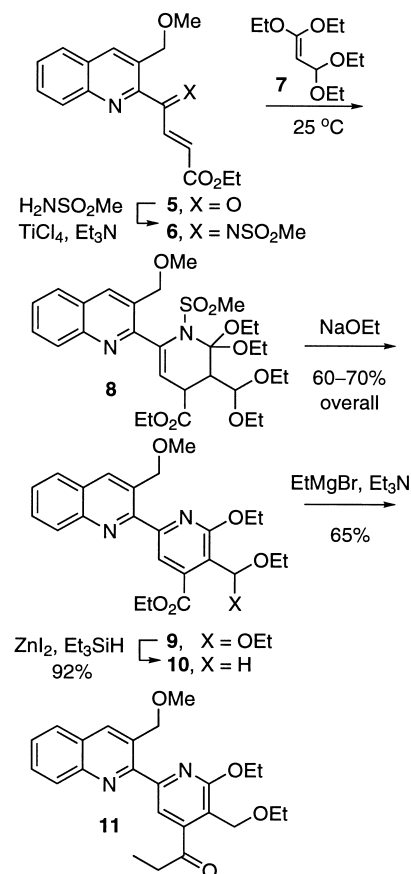
The approach is based on the implementation of a room temperature inverse electron demand Diels–Alder reaction of the *N*-sulfonylimine **6**,²⁷ a *N*-sulfonyl-1-aza-1,3-butadiene,^{28–30} with the electron rich dienophile **7** for introduction of the D-ring pyridone. The azadiene **6** incorporates a noncomplementary C4 electron-withdrawing substituent which is known to lower the diene LUMO accelerating its rate of reaction in an inverse electron demand Diels–Alder reaction without altering the inherent regioselectivity of the cycloaddition.³⁰ Base-catalyzed aromatization of cycloadduct **8** would provide the highly substituted pyridine **9** bearing four of the five pyridone exocyclic substituents, two of which could be further elaborated for introduction of the E-ring. The stereo-



Scheme 1.

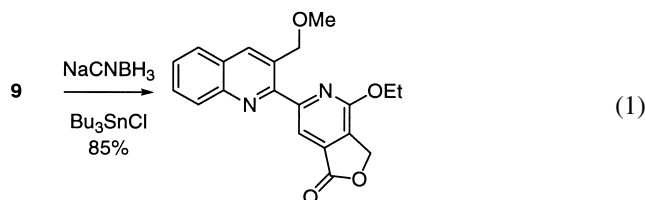
chemistry of the tertiary α -hydroxy group of the E-ring lactone was envisioned to be set by a Sharpless asymmetric dihydroxylation reaction of the corresponding vinyl ether **12**. Finally, the C and E ring closures were expected to come from simultaneous pyridone deprotection and conversion of the benzylic ethers in **17** to the corresponding benzylic bromides, followed by intramolecular displacement upon the addition of base to furnish (+)-camptothecin (Scheme 1).

Methane sulfonamide was condensed with *trans*-ketone **5**²⁷ under dehydrating conditions utilizing TiCl_4 and Et_3N to furnish the *N*-sulfonyl-1-aza-1,3-butadiene **6** as previously detailed, Scheme 2.^{27,31} The inverse electron demand Diels–Alder cycloaddition between the electron rich 1,1,3,3-tetraethoxypropene (**7**)³² and the electron deficient *N*-sulfonyl-1-aza-1,3-butadiene **6** proceeded at room temperature and ambient pressure within 4 h to give the desired [4+2] cycloadduct **8**. Addition of sodium ethoxide to the sensitive Diels–Alder cycloadduct resulted in the sequential elimination of methanesulfinic acid and ethanol to provide the highly substituted 2-ethoxypyridine **9** under mild conditions (0°C , 1–1.5 h). Because of the hydrolysis sensitive nature of both the azadiene **6** and the cycloadduct **8**, the three steps enlisted for the conversion of **6** to pyridine **9** were carried out in higher conversions (60–70% overall) without purification of the intermediates. Due to the labile nature of the diethyl acetal **9**, it was converted to the corresponding ethyl ether using ZnI_2 and Et_3SiH which cleanly gave **10** without any competing lactonization products resulting from capture



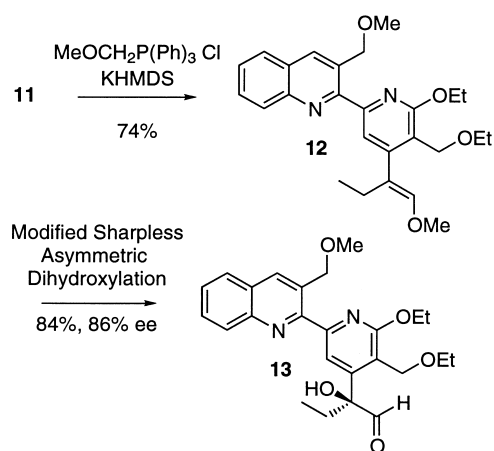
Scheme 2.

of the oxonium intermediate by the adjacent ester. In contrast, treatment of **9** with NaCNBH_3 and Bu_3SnCl ³³ provided such a lactone³⁴ in 85% yield (*t*-BuOH, 80°C, Eq. (1)). Although not extensively investigated, the use of $\text{ZnI}_2/\text{Et}_3\text{SiH}$ was more effective than either $\text{ZnI}_2/\text{NaCNBH}_3$ ³⁵ (20–65%) or $\text{TMSOTf}/\text{Et}_3\text{SiH}$ ³⁶ for conversion of **9** to **10**. In addition, the diethyl acetal of **9** was easily deprotected to provide the corresponding aldehyde³⁷ (TFA, CH_2Cl_2 - H_2O , 0°C, 5–10 min or TsOH, acetone- H_2O , 11 h, quantitative), but alternative routes proceeding through this intermediate have not yet proven competitive.

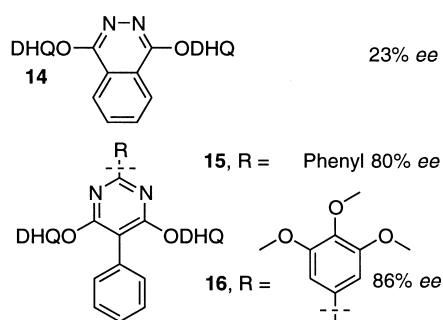


The ethyl ester **10** was directly converted to ethyl ketone **11** by addition of EtMgBr in the presence of excess Et_3N .³⁸ The vinyl ethers **12** were formed through a Wittig reaction of ethyl ketone **11** and methoxymethenetriphenylphosphorane to furnish a 3:1 mixture of *trans/cis* isomers (98%), which could be easily separated providing *E*-**12** (74%), Scheme 3. Analogous to the two additional total syntheses of CPT^{26s,v,y} that utilized a Sharpless asymmetric dihydroxylation,³⁹ the AD reaction of *trans* vinyl ether **12** proved to be difficult initially. Using the commercially available β -AD mix containing ligand **14**, the desired α -hydroxy aldehyde **13** was obtained in low yields and poor enantiomeric excess. Instead of **13** as the major product, ketone **11** was predominately formed resulting from the subsequent oxidative cleavage of the corresponding C–C bond. Reducing the amount of $\text{K}_3\text{Fe}(\text{CN})_6$ to 1.3 equiv. and supplementing the reaction with an additional equivalent at a later time while increasing the amount of ligand **14** and OsO_4 led to an improved yield of the desired product. However, the enantiomeric excess remained low (23% ee).⁴⁰ The enantiomeric excess of the reaction was increased remarkably by utilization of the $(\text{DHQ})_2$ -pyrimidine ligand **15** resulting in 80% ee. The enantioselectivity was further increased to 86% with the 4,6-(DHQ)₂-5-phenyl-1,3-pyrimidine-2-(3,4,5-trimethoxyphenyl) ligand **16** providing material suitable for incorporation into the total synthesis of **1**. When the *cis* vinyl ether was used with the $(\text{DHQD})_2$ version of **16**, **13** was obtained in lower ee's (60–70% ee). Interestingly, when the $(\text{DHQD})_2$ variant of this ligand was used with the *trans* isomer, greater than 99% ee of the corresponding unnatural 20(*R*) enantiomer was obtained.

Once formed, α -hydroxy aldehyde **13** was immediately oxidized to the corresponding carboxylic acid **17** with sodium chlorite⁴¹ in a buffered solution containing resorcinol, Scheme 4. The aryl ethyl ether and the two benzylic ethers were cleaved upon exposure to a saturated solution of $\text{HBr}(\text{g})$ in a 2,2,2-trifluoroethanol at 80°C^{27,29a} to give the corresponding pyridone **18** containing two benzyl bromides. Addition of potassium carbonate to the reaction mixture catalyzed the intramolecular nucleophilic ring closures to give **1**, without detection of competing elimination products.

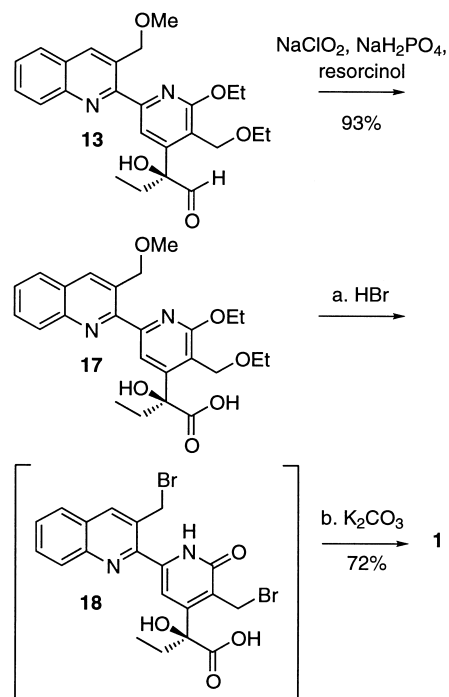


Ligands for Sharpless Asymmetric Dihydroxylation:

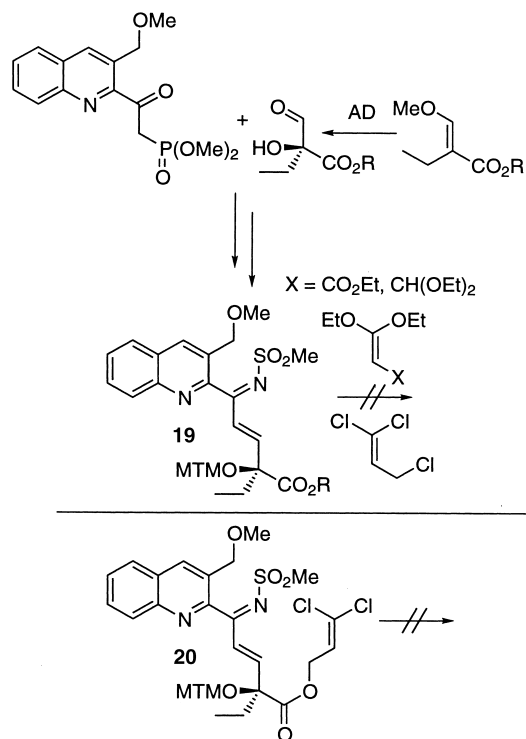


Scheme 3.

Although unanticipated, alternative approaches to camptothecin employing the readily available and more advanced, but more sterically hindered and less electron deficient, *N*-sulfonyl-1-azadiene **19** employing a full range of electron



Scheme 4.



Scheme 5.

rich dienophiles have not proven successful, Scheme 5. Similarly surprising, the intramolecular cycloaddition of the *N*-sulfonyl-1-azadiene **20** was also not successful.

1. Experimental

1.1. General

1.1.1. 3-Diethoxymethyl-2-ethoxy-6-(3-methoxymethylquinolin-2-yl)isonicotinic acid ethyl ester (9). TiCl_4 (1.0 M in CH_2Cl_2 , 4.2 mL) and Et_3N (1.4 mL) was added to a solution of ketone **5**²⁷ (630 mg, 2.1 mmol) and methane sulfonamide (400 mg, 4.2 mmol) in CH_2Cl_2 (63 mL) at -42°C . The brown solution was stirred at -42°C for 40 min, warmed to 25°C , and stirred for an additional 60 min. The crude mixture was poured over a plug of Celite (2.5×7.5 cm) and eluted with a 50% EtOAc–hexane (250 mL). The eluent was concentrated and poured over a plug of silica gel (2.5×7.5 cm) and eluted with 50% EtOAc–hexane (250 mL). The eluent was concentrated and redissolved in benzene (10.5 mL). 1,1,3,3-Tetraethoxypropene (**7**,³² 1.82 g, 8.4 mmol) was added and the solution was stirred for 4 h. The reaction mixture was concentrated in vacuo and the residue was redissolved in THF (21 mL). NaOEt (3.0 M in EtOH, 5.6 mL, 16.8 mmol) was added to the solution at 0°C . The mixture was stirred at 0°C for 75 min and then diluted with EtOAc (50 mL) and H_2O (30 mL). The organic layer was removed and the aqueous layer was washed with EtOAc. The combined organic layers were dried (Na_2SO_4), filtered, and concentrated. Chromatography (SiO_2 , 2.5×20 cm, 5–10% EtOAc–hexane gradient elution) afforded **9** (664 mg, 68%) as a colorless solid: mp 109 – 113°C ; ^1H NMR (CDCl_3 , 600 MHz) δ 8.43 (s, 1H),

8.11 (d, $J=8.3$ Hz, 1H), 7.84 (d, $J=8.3$ Hz, 1H), 7.82 (s, 1H), 7.68 (apparent t, $J=7.5$ Hz, 1H), 7.53 (apparent t, $J=7.5$ Hz, 1H), 5.75 (s, 1H), 5.02 (s, 2H), 4.49 (q, $J=7.0$ Hz, 2H), 4.37 (q, $J=7.0$ Hz, 2H), 3.70 (dq, $J=8.8$, 7.0 Hz, 2H), 3.60 (dq, $J=7.6$, 8.7 Hz, 2H), 3.47 (s, 3H), 1.42 (t, $J=7.0$ Hz, 3H), 1.38 (t, $J=7.0$ Hz, 3H), 1.23 (t, $J=7.0$, 6H); ^{13}C NMR (CDCl_3 , 150 MHz) δ 168.4, 160.1, 155.8, 153.7, 146.5, 143.2, 134.2, 131.1, 129.4, 129.2, 127.8, 127.3, 127.0, 117.6, 115.8, 97.9, 72.1, 63.1 (2C), 62.4, 61.5, 58.6, 15.0 (2C), 14.7, 14.0; IR (KBr) ν_{max} 2978, 2919, 1731, 1560 cm^{-1} ; MALDI-HRMS (dihydroxybenzyl alcohol, DHB) m/z 469.2353 ($\text{M}+\text{H}^+$, $\text{C}_{26}\text{H}_{32}\text{N}_2\text{O}_6$ requires m/z 469.2333).

1.1.2. 2-Ethoxy-3-ethoxymethyl-6-(3-methoxymethylquinolin-2-yl)isonicotinic acid ethyl ester (10). A slurry of ZnI_2 (136 mg, 0.42 mmol), **9** (100 mg, 0.21 mmol), and Et_3SiH (409 μL , 2.56 mmol) in CH_2Cl_2 (750 μL) was stirred for 5 days at 25°C . The slurry was filtered through a plug of silica gel (1×5 cm) and eluted with 30% EtOAc–hexane. Chromatography (SiO_2 , 2.5×10 cm, 20% EtOAc–hexane) afforded **10** (83 mg, 92%) as a colorless solid: mp 75 – 77°C ; ^1H NMR (CDCl_3 , 400 MHz) δ 8.43 (s, 1H), 8.13 (d, $J=8.5$ Hz, 1H), 8.03 (s, 1H), 7.85 (d, $J=8.2$ Hz, 1H), 7.69 (apparent t, $J=7.9$ Hz, 1H), 7.54 (apparent t, $J=7.6$ Hz, 1H), 5.02 (s, 2H), 4.81 (s, 2H), 4.46 (q, $J=7.0$ Hz, 2H), 4.40 (q, $J=7.0$ Hz, 2H), 3.56 (t, $J=7.0$ Hz, 2H), 3.47 (s, 3H), 1.43 (t, $J=7.0$ Hz, 3H), 1.40 (t, $J=7.0$ Hz, 3H), 1.20 (t, $J=7.0$ Hz, 3H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 167.1, 160.8, 155.4, 154.0, 146.5, 142.7, 134.4, 131.1, 129.3, 129.3, 127.8, 127.4, 127.0, 119.2, 116.7, 72.2, 66.2, 63.0, 62.5, 61.5, 58.6, 15.1, 14.7, 14.1; IR (film) ν_{max} 2976, 2868, 1723, 1596, 1562, 1440 cm^{-1} ; MALDI-HRMS (DHB) m/z 425.2071 ($\text{M}+\text{H}^+$, $\text{C}_{24}\text{H}_{28}\text{N}_2\text{O}_5$ requires m/z 425.2071).

1.1.3. 1-[2-Ethoxy-3-ethoxymethyl-6-(3-methoxymethylquinolin-2-yl)pyridin-4-yl]propan-1-one (11). EtMgBr (380 μL , 3.0 M in Et_2O) was added to a solution of ester **10** (220 mg, 0.52 mmol) and Et_3N (2.89 mL, 20.8 mmol) in toluene (10.4 mL) at -5°C and the solution was stirred at -5°C for 5 h. The reaction was quenched by the addition of saturated aqueous NH_4Cl (5 mL). The organic layer was removed and the aqueous layer washed with EtOAc. The combined organic layers were dried (Na_2SO_4), filtered, and concentrated. PTLC (SiO_2 , 20% EtOAc–hexane) afforded **11** (141 mg, 66%) as a colorless solid: mp 102 – 105°C ; ^1H NMR (CDCl_3 , 500 MHz) δ 8.44 (s, 1H), 8.12 (d, $J=8.5$ Hz, 1H), 7.85 (d, $J=8.1$ Hz, 1H), 7.74 (s, 1H), 7.69 (dt, $J=8.1$, 8.4 Hz, 1H), 7.54 (dt, $J=8.1$, 8.1 Hz, 1H), 5.04 (s, 2H), 4.65 (s, 2H), 4.45 (q, $J=7.4$ Hz, 2H), 3.52 (q, $J=7.0$ Hz, 2H), 3.48 (s, 3H), 2.92 (q, $J=7.3$ Hz, 2H), 1.42 (t, $J=7.4$ Hz, 3H), 1.22 (t, $J=7.4$ Hz, 3H), 1.19 (t, $J=7.0$ Hz, 3H); ^{13}C NMR (CDCl_3 , 125 MHz) δ 206.0, 160.1, 155.2, 154.0, 151.5, 146.5, 134.4, 131.2, 129.3 (2C), 127.8, 127.4, 127.0, 117.0, 114.3, 72.3, 66.6, 63.7, 62.4, 58.6, 36.1, 15.0, 14.7, 7.6; IR (film) 2970, 2860, 1701, 1596, 1555, 1437, 1378, 1331 cm^{-1} ; MALDI-HRMS (DHB) m/z 409.2134 ($\text{M}+\text{H}^+$, $\text{C}_{24}\text{H}_{28}\text{N}_2\text{O}_4$ requires m/z 409.2122).

1.1.4. 2-[6-Ethoxy-5-ethoxymethyl-4-(1-methoxymethylenepropyl)pyridin-2-yl]-3-methoxymethylquinoline (12). KHMDs (0.5 M in toluene, 6.61 mL) was added to a

slurry of methoxymethyltriphenylphosphonium chloride (1.18 g, 3.31 mmol) in THF (10 mL) at 0°C. The red solution was stirred at 0°C for 30 min before ketone **11** (270 mg, 0.66 mmol) was added in THF (1 mL). The solution was warmed to 25°C and stirred for 10 h. Saturated aqueous NH₄Cl (5 mL) was added and the organic layer removed. The aqueous layer was washed with EtOAc and the combined organic layers were dried (Na₂SO₄), filtered, and concentrated. Chromatography (SiO₂, 2.5×10 cm, 5–10% EtOAc–hexane gradient elution) afforded a high R_f oil (213 mg, 74%) and a lower R_f oil (76 mg, 26%).

High R_f product (*trans*-**12**): ¹H NMR (CDCl₃, 400 MHz) δ 8.42 (s, 1H), 8.12 (d, *J*=8.5 Hz, 1H), 7.84 (d, *J*=8.2 Hz, 1H), 7.68 (apparent t, *J*=7.6 Hz, 1H), 7.63 (s, 1H), 7.52 (apparent t, *J*=7.6 Hz, 1H), 6.35 (s, 1H), 5.03 (s, 2H), 4.48 (s, 2H), 4.44 (q, *J*=7.3 Hz, 2H), 3.70 (s, 3H), 3.65 (q, *J*=6.8 Hz, 2H), 3.48 (s, 3H), 2.55 (q, *J*=7.6 Hz, 2H), 1.43 (t, *J*=6.8 Hz, 3H), 1.28 (t, *J*=7.0 Hz, 3H), 0.96 (t, *J*=7.3 Hz, 3H); ¹³C NMR (CDCl₃, 150 MHz) δ 162.1, 155.1, 154.4, 152.8, 147.5, 146.5, 134.0, 131.4, 129.3, 129.0, 127.7, 127.4, 126.7, 118.2, 117.8, 117.7, 72.3, 65.8, 64.8, 62.0, 59.9, 58.6, 21.8, 15.4, 14.8, 12.9; IR (film) 2968, 2926, 2864, 1651, 1594, 1547, 1444, 1372, 1330 cm⁻¹; MALDI-HRMS (DHB) *m/z* 437.2435 (M+H⁺, C₂₆H₃₂N₂O₄ requires *m/z* 437.2435).

Low R_f product (*cis*-**12**): ¹H NMR (CDCl₃, 400 MHz) δ 8.43 (s, 1H), 8.11 (d, *J*=8.5 Hz, 1H), 7.84 (d, *J*=8.2 Hz, 1H), 7.61 (apparent t, *J*=8.2 Hz, 1H), 7.61 (s, 1H), 7.51 (apparent t, *J*=8.2 Hz, 1H), 6.01 (s, 1H), 5.08 (s, 2H), 4.51 (s, 2H), 4.45 (q, *J*=7.0 Hz, 2H), 3.57 (q, *J*=7.0 Hz, 2H), 3.54 (s, 3H), 3.50 (s, 3H), 2.33 (q, *J*=7.3 Hz, 2H), 1.43 (t, *J*=7.0 Hz, 3H), 1.22 (t, *J*=7.0 Hz, 3H), 1.03 (t, *J*=7.4 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 161.6, 154.9, 154.5, 151.0, 146.5, 142.3, 133.8, 131.4, 129.3, 129.0, 127.7, 127.4, 126.6, 118.5, 118.3, 117.5, 72.4, 65.9, 64.4, 61.9, 59.6, 58.6, 26.8, 15.3, 14.8, 13.3; IR (film) 2968, 2919, 2860, 1661, 1590, 1548, 1437, 1378, 1331 cm⁻¹; MALDI-HRMS (DHB) *m/z* 437.2423 (M+H⁺, C₂₆H₃₂N₂O₄ requires *m/z* 437.2435).

1.1.5. 2-[2-Ethoxy-3-ethoxymethyl-6-(3-methoxymethyl-quinolin-2-yl)pyridin-4-yl]-2-hydroxybutyraldehyde (13). A slurry of (DHQ)₂-Pyr(OMe)₃ (**16**, 4.1 mg, 0.0047 mmol), K₂CO₃ (12.3 mg, 0.09 mmol), K₃Fe(CN)₆, and OsO₄ (23 μL of 4% wt in H₂O, 0.0035 mmol) in H₂O (300 μL) was stirred at 0°C for 5 min before *trans*-**12** (13 mg, 0.030 mmol) in *t*-BuOH (300 μL) was added. After 4 h at 0°C, additional K₃Fe(CN)₆ (10 mg, 0.03 mmol) was added and the solution was stirred at 0°C for 4 h. Solid sodium sulfite (40 mg, 0.3 mmol) was added and the solution stirred for 30 min at 0°C. CH₂Cl₂ (600 μL) was added and the organic layer was removed. The aqueous layer was extracted (3×600 μL) with CH₂Cl₂ and the combined organic layers were passed through a small plug of silica gel (1×4 cm) and eluted with 50% EtOAc–hexane. The eluent was concentrated by a steady stream of N₂. Chromatography (SiO₂, 20% EtOAc–hexane) afforded **13** (11 mg, 84%) as a colorless oil: [α]_D²⁵=+20 (c 0.011, CH₂Cl₂); ¹H NMR (CDCl₃, 600 MHz) δ 9.74 (s, 1H), 8.44 (s, 1H), 8.13 (d, *J*=8.3 Hz, 1H), 7.90 (s, 1H), 7.86 (d, *J*=7.9 Hz, 1H), 7.70 (apparent t,

J=7.0 Hz, 1H), 7.55 (apparent t, *J*=7.0 Hz, 1H), 5.49 (br s, 1H), 5.02 (s, 2H), 4.89 (d, *J*=11.4 Hz, 1H), 4.65 (d, *J*=11.4 Hz, 1H), 4.44 (q, *J*=7.0 Hz, 2H), 3.60 (q, *J*=7.0 Hz, 2H), 3.48 (s, 3H), 2.27 (m, 2H), 1.42 (t, *J*=7.0 Hz, 3H), 1.27 (t, *J*=7.0 Hz, 3H), 1.01 (t, *J*=7.0 Hz, 3H); ¹³C NMR (CDCl₃, 150 MHz) δ 200.6, 161.6, 155.3, 154.3, 150.7, 146.6, 134.4, 131.2, 129.5, 129.3, 127.8, 127.4, 127.0, 117.9, 116.4, 82.7, 72.3, 66.2, 63.5, 62.6, 58.7, 29.6, 14.9, 14.7, 7.4; IR (film) 3389, 2919, 2848, 1731, 1549, 1378, 1331 cm⁻¹; MALDI-HRMS (DHB) *m/z* 439.2219 (M+H⁺, C₂₅H₃₀N₂O₅ requires *m/z* 439.2227).

1.1.6. 2-[2-Ethoxy-3-ethoxymethyl-6-(3-methoxymethyl-quinolin-2-yl)pyridin-4-yl]-2-hydroxybutyric acid (17). Aldehyde **13** (10 mg, 0.023 mmol) and resorcinol (25 mg, 0.23 mmol) were dissolved in DMSO (600 μL). NaH₂PO₄ (18 mg, 0.115 mmol) in H₂O (200 μL) was added and the mixture cooled to 0°C before NaClO₂ (10.3 mg, 0.115 mmol) was added. The purple mixture was stirred for 1 h at 0°C before saturated aqueous NH₄Cl (1 mL) was added. The solution was extracted with EtOAc (4×500 μL) and the combined organic layers were dried (NaSO₄), filtered, and concentrated. Chromatography (SiO₂, 50–100% EtOAc–hexane, followed by 15% MeOH–CH₂Cl₂) afforded **17** (9.6 mg, 93%) as an amorphous solid: [α]_D²⁵=+29 (c 0.010, MeOH); ¹H NMR (*d*₆-DMSO, 400 MHz) δ 8.49 (s, 1H), 8.12 (s, 1H), 8.08 (d, *J*=8.2 Hz, 1H), 8.05 (d, *J*=7.6 Hz, 1H), 7.75 (apparent t, *J*=7.0 Hz, 1H), 7.62 (t, *J*=7.0 Hz, 1H), 4.99 (m, 3H), 4.69 (d, *J*=10.0 Hz, 1H), 4.38 (q, *J*=7.0 Hz, 2H), 3.53 (q, *J*=7.0 Hz, 2H), 3.41 (s, 3H), 3.25–3.40 (br s, 1H), 2.04 (m, 2H), 1.37 (t, *J*=7.0 Hz, 3H), 1.14 (t, *J*=6.8 Hz, 3H), 0.82 (t, *J*=7.0 Hz, 3H); ¹³C NMR (*d*₆-DMSO, 100 MHz) δ 174.5, 161.9, 154.9, 152.8, 145.9, 134.1, 131.1, 129.5, 128.7, 127.7, 127.1, 126.9 (2C), 118.8, 116.2, 79.1, 71.6, 65.3, 63.3, 61.5, 58.1, 32.2, 15.3, 14.8, 8.8; IR (film) 3334, 2974, 2922, 1644, 1691, 1510, 1592, 1548, 1372, 1333 cm⁻¹; MALDI-HRMS (DHB) *m/z* 455.2172 (M+H⁺, C₂₅H₃₀N₂O₆ requires *m/z* 455.2177).

1.1.7. (+)-Camptothecin (1). A sample of **17** (9.4 mg, 0.021 mmol) was added to a freshly prepared solution of concentrated HBr(g) in 2,2,2-trifluoroethanol (1.88 mL). The mixture was warmed at reflux for 24 h in a sealed vial, cooled to 25°C, and stirred for an additional 14 h. K₂CO₃ (14 mg, 0.1 mmol) was added to the solution and the mixture was stirred for 1 h. The mixture was filtered through a plug of silica gel (1×5 cm) and eluted with 10% MeOH–CH₂Cl₂. The eluent was concentrated and purified by chromatography (SiO₂, 0–10% MeOH–CH₂Cl₂) to afford **1** (5.2 mg, 72%) as an off-white solid identical in all respects with authentic material: [α]_D²⁵=+30 (c 0.001, MeOH–CH₂Cl₂, 1:4), lit [α]_D²⁵=+32 (c 0.4, MeOH–CH₂Cl₂, 1:4); ¹H NMR (CD₂Cl₂/CD₃OD, 600 MHz) δ 8.48 (s, 1H), 8.18 (d, *J*=8.3 Hz, 1H), 7.98 (d, *J*=7.9 Hz, 1H), 7.82 (apparent t, *J*=7.0 Hz, 1H), 7.67 (t, *J*=7.0 Hz, 1H), 7.67 (s, 1H), 5.63 (d, *J*=16.2 Hz, 1H), 5.30 (d, *J*=16.2 Hz, 1H), 5.28 (s, 2H), 1.90 (m, 2H), 0.99 (t, *J*=7.4 Hz, 3H); ¹³C NMR (CD₂Cl₂/CD₃OD, 150 MHz) δ 174.1, 158.4, 152.8, 151.5, 149.0, 146.5, 132.2, 131.2, 129.5, 129.4, 128.8, 128.8, 1128.5, 119.6, 99.0, 73.3, 66.4, 50.7, 31.7, 7.8; MALDI-HRMS (DHB) *m/z* 349.1185 (M+H⁺, C₂₀H₁₆N₂O₄ requires *m/z* 349.1183).

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